

Metabolism of C₂₈ Phytosterols in the Insect *Tenebrio Molitor*: Migration of the C-25 Hydrogen Atom to C-24

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During the conversion of 24-methylenecholesterol into cholesterol by *Tenebrio molitor* larvae the C-25 hydrogen migrates to the C-24 position.

Cholesterol can be obtained by many phytophagous insects through the dealkylation of C₂₈ and C₂₉ phytosterols that they find in their diet.¹ The mechanism of this process has been thoroughly investigated for C₂₉ phytosterols for which the intermediacy of the stereoisomeric 24,28-ethylidene compounds (**2a**) (Scheme 1)² and 24,28-epoxides (**3a**)³ has been demonstrated. A peculiar feature in the above metabolic sequence is the hydrogen migration from the C-25 to the C-24 position during the conversion of the 24,28-epoxides (**3a**) into cholesterol (**4**)⁴ as illustrated in Scheme 1.

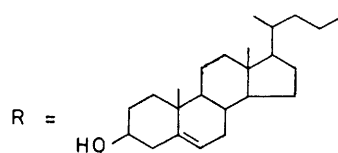
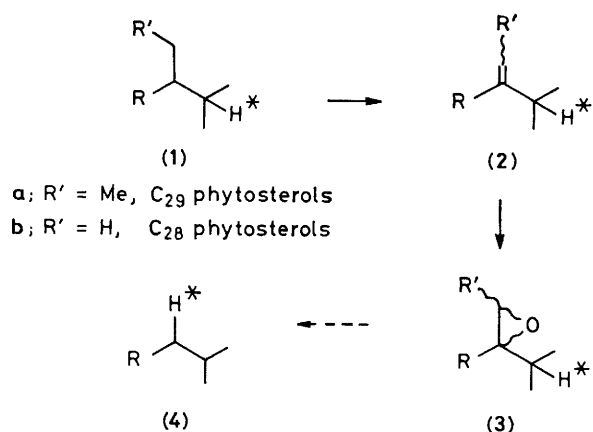
We have recently observed⁵ that the dealkylation of C₂₈ phytosterols seems to proceed in a similar way: in fact both 24-methylenecholesterol (**2b**) and the corresponding 24,28-epoxides (**3b**) are converted into cholesterol (**4**) by *Tenebrio molitor* larvae. Moreover, 24-methylenecholesterol epoxide has been shown to be an intermediate in the formation of cholesterol in *Schistocerca* mid-gut microsomes.⁶

We now report our results on the mechanism of this process, *i.e.* the demonstration that the hydrogen shift from the C-25 to the C-24 position occurs also in the C₂₈ phytosterol dealkylation.

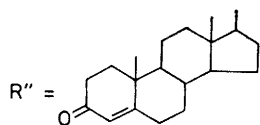
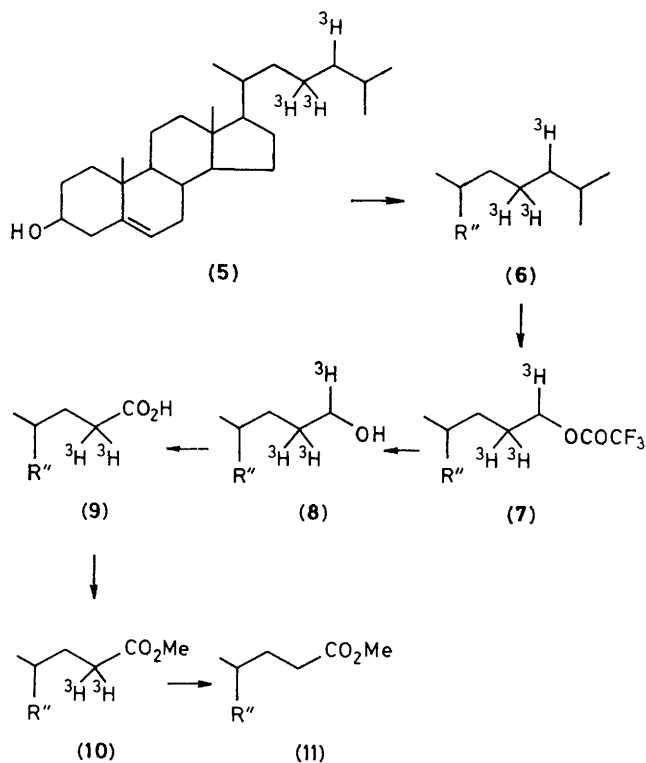
A mixture of [23,23,25-³H₃]-24-methylenecholesterol (5.96

× 10⁷ dpm of ³H, spec. act. 2.64 × 10⁷ dpm/mg), synthesized from [23,23,25-³H₃]-24-oxocholesterol,⁷ and [4-¹⁴C]sitosterol (7.86 × 10⁶ dpm of ¹⁴C, spec. act. 2.85 × 10⁸ dpm/mg, the Radiochemical Centre, Amersham) as internal standard, was fed to 500 young *Tenebrio molitor* larvae. After four days the larvae were sacrificed and the unsaponifiable fraction, obtained as previously described,³ was benzoylated. Cholesteryl benzoate was obtained pure by sequential argentation t.l.c. (to remove the unconverted 24-methylenecholesteryl benzoate) and preparative h.p.l.c. (Waters μBondapack-C₁₈ column, flow rate 2.5 ml/min, solvent MeOH-H₂O 97:3), which removed the residual sitosteryl benzoate.

The labelled cholesteryl benzoate, diluted with cold material, was subjected to alkaline hydrolysis and to Oppenauer oxidation to yield labelled cholest-4-en-3-one (**6**) (Scheme 2). Trifluoroacetic acid oxidation of (**6**)⁸ gave 24-trifluoroacetoxychol-4-en-3-one (**7**) with loss of the terminal isopropyl group; hydrolysis to the 24-hydroxy-compound (**8**) followed by oxidation with pyridinium dichromate (PDC)⁹ afforded the 24-carboxy-derivative (**9**); esterification with diazomethane and exchange of the obtained methyl 3-oxochol-4-en-24-oate (**10**) with MeOH/MeO⁻ gave the methyl ester



Scheme 1



Scheme 2

Table 1. Specific activities and ³H/¹⁴C ratios of cholesterol and its chemical degradation products.

Compounds	dpm of ¹⁴ C/mmol	³ H/ ¹⁴ C ^c
(5)	1.13 × 10 ⁵	13.6
(6)	1.14 × 10 ⁵	13.7
(8)	1.15 × 10 ⁵	13.2
(10) ^a	1.08 × 10 ⁵	8.2
(11) ^b	1.10 × 10 ⁵	0.9

^a Methyl 3-oxochole-4-en-24-oate before MeOH/MeO⁻ exchange.^b Methyl 3-oxochole-4-en-24-oate after MeOH/MeO⁻ exchange.^c Ratio of dpm of ³H/mmol to dpm of ¹⁴C/mmol.

(11) with almost complete loss of tritium.

Each product of the above sequence was crystallized to constant specific activity and ³H/¹⁴C ratio and the values obtained are reported in Table 1.

The total retention of the tritium label during the peracetic acid oxidation of (6) shows the absence of label at C-25; moreover the loss of 40% of tritium during the PDC oxidation of (8) is indicative of the presence of tritium at C-24. The virtually complete loss of the remaining tritium by MeOH/MeO⁻ exchange of (10) indicates that this residual tritium is present at C-23.

These results clearly demonstrate that during the metabolism of 24-methylenecholesterol by *Tenebrio molitor*, the C-25 hydrogen migrates to the C-24 position and they also provide further evidence for the strict similarity of the de-ethylation process of C₂₉ phytosterols and the demethylation process of their C₂₈ analogues.†

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† A recent communication (S. Maruyama, Y. Fujimoto, M. Morisaki, and N. Ikekawa, *Tetrahedron Lett.*, 1982, **23**, 1701) actually confirms 24-methylenecholesterol as an intermediate and suggests a possible migration of the hydrogen from the C-25 to the C-24 position during the demethylation of campesterol (1b) in the insect *Bombyx mori*.